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Tetrahedron Letters 41 (2000) 3119–3122

TETRAHEDRON  
LETTERS

## Efficient synthesis of L-altrose and L-mannose

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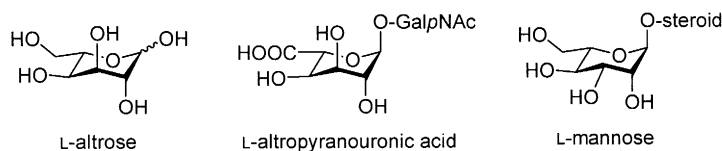
Received 20 December 1999; revised 25 January 2000; accepted 25 February 2000

### Abstract

Two convenient routes for the synthesis of L-altrose and L-mannose from 1,2:3,5-di-*O*-isopropylidene- $\beta$ -L-idofuranose in four and six steps via the stereoselective hydroboration and hydrogenation of olefins as key steps are described here, respectively. © 2000 Elsevier Science Ltd. All rights reserved.

**Keywords:** carbohydrates; glycosides; L-altrose; L-mannose.

L-Altropyranosyl and L-mannopyranosyl units are structural elements of several naturally occurring products, as outlined in Scheme 1. L-Altrose is a component of the extracellular polysaccharide from *Butyrivibrio fibrisolvens* strain CF3.<sup>1</sup> The O-specific polysaccharide of the bacterium *Proteus mirabilis*, a human opportunistic pathogen causing urinary tract infections, contain L-altropyranouronic acid as a new component of O-antigens.<sup>2</sup> L-Mannopyranosides had been found in the sugar units of steroid glycosides<sup>3</sup> and their phenol derivatives are potent substrates for determining the  $\alpha$ -L-mannosidase activity of commercial naringinase.<sup>4</sup>

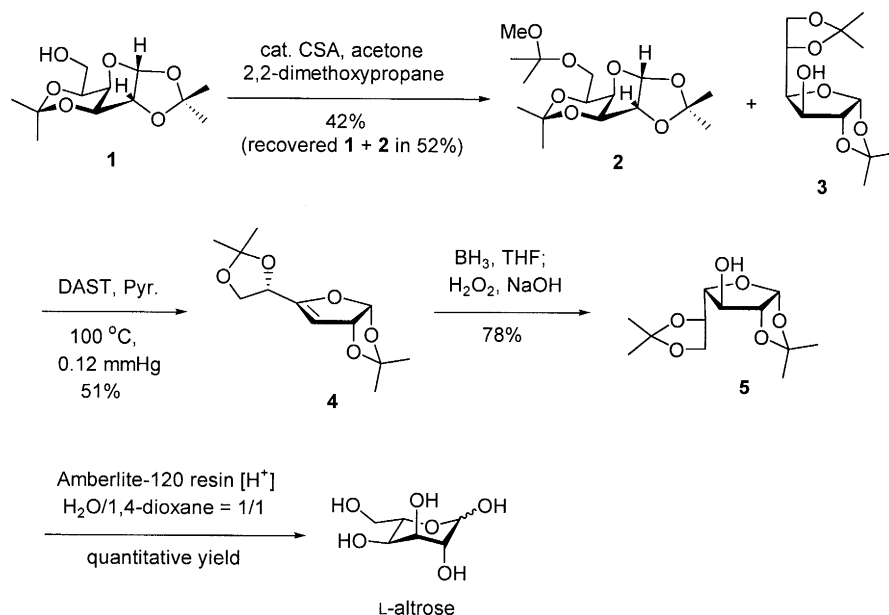


Scheme 1.

Approaches for the synthesis of L-altrose have been reported including the asymmetric Sharpless epoxidation of olefins followed by ring opening,<sup>5</sup> intramolecular Tishchenko reactions of protected hexos-5-uloses<sup>6</sup> and bromination of  $\Delta^4$ -uronate followed by a three-step conversion.<sup>7</sup> Some efforts had also been made toward the L-mannose comprising the asymmetric hetero Diels–Alder reaction followed by stereoselective hydroboration<sup>8</sup> and the applications of enantioselective Sharpless epoxidation<sup>5</sup> as well as dihydroxylation<sup>9</sup> of olefins. In association with our interest in the synthesis of biologically important L-hexoses, we have explored herein two short routes for the synthesis of L-altrose and L-mannose, respectively.

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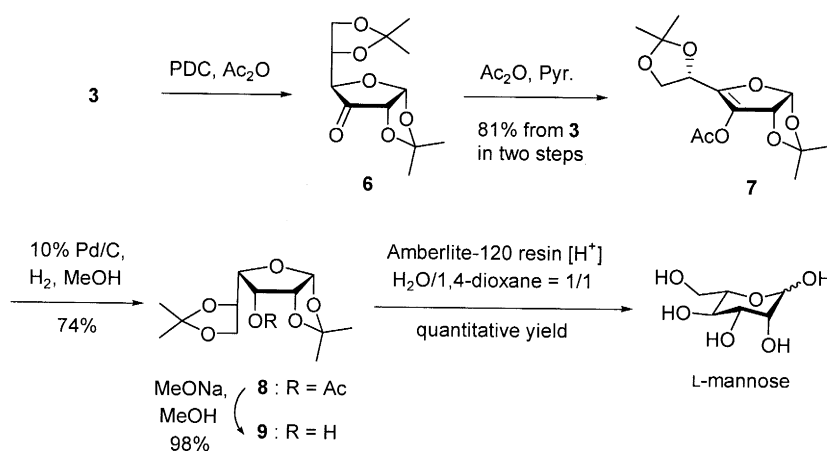
Scheme 2 illustrates an efficient synthesis of L-altrose from 1,2:3,5-di-*O*-isopropylidene- $\beta$ -L-idofuranose **1** in four steps. The starting material **1**, generated from diacetone  $\alpha$ -D-glucose in three steps,<sup>10</sup> underwent orthogonal isopropylidene rearrangement with a solution of 2,2-dimethoxypropane and acetone in the presence of catalytic amount of ( $\pm$ )-camphorsulfonic acid at room temperature to provide 1,2:5,6-di-*O*-isopropylidene- $\beta$ -L-idofuranose **3** as a white solid (42% after recrystallization from hexane) and a mixture of **2** and unreacted **1** in ca. 52% yield. This mixture can be re-utilized under the same conditions and similar results are obtained. Epimerisation of **3** at C-4 into 1,2:5,6-di-*O*-isopropylidene- $\beta$ -L-altrofuranose **5** could be carried out by sequential elimination of H<sub>2</sub>O and stereoselective hydroboration via the olefin **4** as an intermediate. Diethylaminosulfur trifluoride (DAST)<sup>11</sup> and pyridine were consecutively added to a solution of **3** in dichloromethane at 0°C under nitrogen. The mixture was subjected to distillation by the Kugelrohr apparatus at 100°C under 0.12 mmHg to give the olefin **4**<sup>12</sup> (51%) which was treated with borane reagent followed by oxidative work-up to provide **5**<sup>12</sup> as a single isomer in 78% yield. The high stereoselectivity is perhaps due to the steric hindrance of a 5,5-*cis*-fused ring configuration, forming the 3-hydroxy group toward up-face. Acidic hydrolysis of **5** in the presence of Amberlite-120 resin (H<sup>+</sup> form) afforded the desired L-altrose in quantitative yield. Comparison of our data with the literature report<sup>13</sup> revealed identity with respect to <sup>1</sup>H and <sup>13</sup>C NMR spectra.



Scheme 2.

The preparation of L-mannose from **3** in five steps is summarized in Scheme 3. The key step involving the inversion of chiral centers in **3** at C-3 and C-4 positions could be achieved by *syn*-reduction of the enol acetate **7**. Oxidation of **3** with PDC and acetic anhydride<sup>14</sup> led to the ketone **6** which underwent enolization in pyridine and acetic anhydride to furnish the enol acetate **7**<sup>12</sup> (81% from **3** in two steps). Stereoselective hydrogenation in the presence of 10% Pd/C as the catalyst provided 3-*O*-acetyl-1,2:5,6-di-*O*-isopropylidene- $\beta$ -L-mannofuranose **8**<sup>12</sup> in 74% yield. The highly stereoselective *syn*-addition of hydrogen molecule with 3-*endo* double bond occurring from less-hindered side is perhaps induced by the *cis*-fused ring junction. Similar transformation of 3-*O*-acetyl-1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-*erythro*-hex-3-endofuranose into 1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-gulofuranose via the NaBH<sub>4</sub> reduction had

been reported in 34% yield.<sup>15</sup> The yield can be improved by the hydrogenation reaction. Deacetylation of **8** with sodium methoxide in methanol obtained **9** (98%) which was hydrolyzed in acidic media to afford the target L-mannose in quantitative yield. The <sup>1</sup>H and <sup>13</sup>C NMR spectra are identical with the literature report.<sup>16</sup>



Scheme 3.

In conclusion, two convenient routes for the synthesis of L-altrose, L-mannose and their furanosyl derivatives are successfully developed here, respectively. The application of these carbohydrates for the synthesis of biologically important oligosaccharides is currently under investigation.

## Acknowledgements

We thank Prof. Sunney I. Chan for his helpful discussions and the National Science Council of Republic of China for financial support.

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- The selected physical data of key compounds is listed. Compound **4**: IR (CHCl<sub>3</sub>)  $\nu$  1669 (m) cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.05 (d, *J*=5.1 Hz, 1H, H-1), 4.27 (dd, *J*=5.1, 2.2 Hz, 1H, H-2), 5.23 (d, *J*=2.2 Hz, 1H, H-3), 4.57 (t, *J*=6.8 Hz, 1H, H-5), 4.12 (dd, *J*=8.3, 6.8 Hz, 1H, H-6), 3.91 (dd, *J*=8.3, 6.8 Hz, 1H, H-6), 1.43 (s, 3H, C-CH<sub>3</sub>), 1.41 (s, 3H, C-CH<sub>3</sub>), 1.39 (s, 3H, C-CH<sub>3</sub>), 1.37 (s, 3H, C-CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  159.76 (C), 112.26 (C), 110.29 (C), 106.50 (CH), 99.42 (CH), 83.33 (CH), 71.44 (CH), 67.21 (CH<sub>2</sub>), 28.12 (CH<sub>3</sub>), 27.81 (CH<sub>3</sub>), 26.09 (CH<sub>3</sub>), 25.48 (CH<sub>3</sub>). Compound **5**:  $[\alpha]_D^{25} = -17.7$  (c 1.0, CHCl<sub>3</sub>); mp=87–88°C; IR (CHCl<sub>3</sub>)  $\nu$  3390 (s) cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  5.85 (d, *J*=3.9

Hz, 1H, H-1), 4.54 (d,  $J=3.9$  Hz, 1H, H-2), 4.40 (t,  $J=2.5$  Hz, 1H, H-3), 4.23 (ddd,  $J=9.4, 6.1, 5.0$  Hz, 1H, H-5), 4.09 (dd,  $J=8.7, 6.1$  Hz, 1H, H-6), 3.92 (dd,  $J=8.7, 5.0$  Hz, 1H, H-6), 3.77 (dd,  $J=9.4, 2.5$  Hz, 1H, H-4), 2.96 (d,  $J=3.9$  Hz, 1H, OH), 1.49 (s, 3H, C-CH<sub>3</sub>), 1.41 (s, 3H, C-CH<sub>3</sub>), 1.33 (s, 3H, C-CH<sub>3</sub>), 1.30 (s, 3H, C-CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  112.76 (C), 109.71 (C), 105.59 (CH), 88.27 (CH), 87.05 (CH), 76.68 (CH), 75.56 (CH), 67.62 (CH<sub>2</sub>), 27.04 (CH<sub>3</sub>), 26.90 (CH<sub>3</sub>), 26.10 (CH<sub>3</sub>), 25.31 (CH<sub>3</sub>). Anal. calcd for C<sub>12</sub>H<sub>20</sub>O<sub>6</sub>: C,55.37;H,7.74, found: C,55.68;H,7.79. Compound 7: [ $\alpha$ ]<sub>D</sub><sup>28</sup> = -41.5 (c 1.0, CHCl<sub>3</sub>); IR (neat)  $\nu$  1751 (s), 1623 (m) cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  6.02 (d,  $J=5.4$  Hz, 1H, H-1), 5.38 (d,  $J=5.4$  Hz, 1H, H-2), 4.67 (t,  $J=7.0$  Hz, 1H, H-5), 4.10 (dd,  $J=8.3, 7.0$  Hz, 1H, H-6), 3.99 (dd,  $J=8.3, 7.0$  Hz, 1H, H-6), 2.19 (s, 3H), 1.46 (s, 3H), 1.43(s, 3H), 1.43(s, 3H), 1.37(s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  168.42 (C), 144.85 (C), 128.94 (C), 113.27 (C), 110.39 (C), 103.86 (CH), 81.03 (CH), 68.92 (CH), 65.73 (CH<sub>2</sub>), 27.88 (CH<sub>3</sub>), 27.80 (CH<sub>3</sub>), 25.68 (CH<sub>3</sub>), 20.48 (CH<sub>3</sub>). Compound 8: [ $\alpha$ ]<sub>D</sub><sup>30</sup> = +26.6 (c 1.0, CHCl<sub>3</sub>); IR (neat)  $\nu$  1750 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  5.70 (d,  $J=4.1$  Hz, 1H, H-1), 5.16 (t,  $J=5.8$  Hz, 1H, H-3), 4.74 (dd,  $J=5.8, 4.1$  Hz, 1H, H-2), 4.50 (dt,  $J=8.7, 6.0$  Hz, 1H, H-5), 4.07 (dd,  $J=8.7, 6.0$  Hz, 1H, H-6), 4.01 (dd,  $J=8.7, 6.0$  Hz, 1H, H-4), 3.97 (dd,  $J=8.7, 6.0$  Hz, 1H, H-6), 2.12 (s, 3H), 1.54 (s, 3H), 1.34 (s, 3H), 1.31 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  170.15 (C), 114.63 (C), 109.57 (C), 104.68 (CH), 79.44 (CH), 73.49 (CH), 71.11 (CH), 67.18 (CH<sub>2</sub>), 26.90 (CH<sub>3</sub>), 26.73 (CH<sub>3</sub>), 25.56 (CH<sub>3</sub>), 20.74 (CH<sub>3</sub>). Compound 9: [ $\alpha$ ]<sub>D</sub><sup>30</sup> = +3.4 (c 1.1, CHCl<sub>3</sub>); IR (neat)  $\nu$  3502 (br) cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  5.68 (d,  $J=4.2$  Hz, 1H, H-1), 4.65 (dd,  $J=5.8, 4.2$  Hz, 1H, H-2), 4.55 (dt,  $J=8.7, 6.0$  Hz, 1H, H-5), 4.28 (t,  $J=5.8$  Hz, 1H, H-3), 4.11 (dd,  $J=8.7, 6.0$  Hz, 1H, H-6), 3.96 (dd,  $J=8.7, 6.0$  Hz, 1H, H-6), 3.83 (dd,  $J=8.7, 5.8$  Hz, 1H, H-4), 3.00–2.80 (bs, 1H, OH) 1.57 (s, 3H), 1.41 (s, 3H), 1.38 (s, 3H), 1.35 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  114.77 (C), 109.67 (C), 105.06 (CH), 81.87 (CH), 80.13 (CH), 73.56 (CH), 70.06 (CH), 67.38 (CH<sub>2</sub>), 26.99 (CH<sub>3</sub>), 26.79 (CH<sub>3</sub>), 25.37 (CH<sub>3</sub>).

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